DRB5*01:08N Product Insert

101.812-12 – including *Taq* **polymerase**, IFU-01 **101.812-12u – without** *Taq* **polymerase**, IFU-02

Visit https://labproducts.caredx.com for "Instructions for Use" (IFU)

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Lot No.: **5F6** Lot-specific information

Olerup SSP® DRB5*01:08N

Product number: 101.812-12 – including *Taq* polymerase

101.812-12u - without *Taq* polymerase

Lot number: 5F6

Expiry date: 2021-09-01

Number of tests: 12 Number of wells per test: 2+1

Storage - pre-aliquoted primers: dark at -20°C

PCR Master Mix: -20°C
Adhesive PCR seals
Product Insert
RT

This Product Description is only valid for Lot No. 5F6.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

Changes compared to the previous *OLERUP* SSP® DRB5*01:08N Lot (2F5)

The DRB5*01:08N specificity and interpretation tables have been updated compared the previous *Olerup* SSP® DRB5*01:08N lot (**Lot No. 2F5**). The kit design is based on IMGT/HLA database 3.28.0.

The DRB5*01:08N primer set is unchanged compared to the previous lot.

Changes in revision R01 compared to R00:

1. The expiration date has been altered due to extension of shelf-life.

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Well **3** contains <u>Negative Control primer pairs</u>, that will amplify more than 95% of the *Olerup* SSP® HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs. The PCR product generated by the positive control primer pair is 430 base pairs.

Length of PCR	105	200	105	80	75	80	85
product							
5'-primer ¹	164	340	440	45	45	43	36
•	5'-CAC3'	^{5'} -Agg ^{3'}	^{5'} -TTA3'	⁵ '-Tgg ³ '	⁵ '-Tgg ³ '	⁵ '-Tgg ³ '	^{5'} -TAC ^{3'}
							36
							^{5'} -TAT ^{3'}
3'-primer ²	231	2 nd I	507	59	58	57	47
	⁵ '-TgC ³ '	^{5'} -AAA ^{3'}	^{5'} -TTg ^{3'}	5'-CTC3'	^{5'} -ggC ^{3'}	5'-CTC3'	5'-ACA3'
							48
							^{5'} -gCA ^{3'}
							48
							^{5'} -gCC ^{3'}
							52
							^{5'} -TgT ^{3'}
A*	+	+	+				
B*	+	+	+				
C*	+	+	+				
DRB1				+	+		
DRB3				+	+		
DRB5				+			
DQB1					+		
DPB1						+	
DQA1							+

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codonnumbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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PRODUCT DESCRIPTION

DRB5*01:08N SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DRB5*01:08N allele.

PLATE LAYOUT

Each test consists of 3 PCR reactions in an 8 well cut PCR plate. Wells 4 to 8 are empty.

1 2 NC empty empty empty empty empty

The 8 well cut PCR plate is marked with the Lot No. '5F6' in silver/gray ink.

Well No. 1 is marked with the Lot No. '5F6'.

Wells 1 to 2 – DRB5*01:08N high resolution primers.

Well 3 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded. The PCR plates are heat-sealed with a PCR-compatible foil.

Please note: When removing each 8 well PCR plate, make sure that the remaining plates stay sealed. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

The interpretation of DRB5*01:08N SSP subtypings will be influenced by the other DRB5 alleles.

UNIQUELY IDENTIFIED ALLELES

The DRB5*01:08N allele will give rise to a unique amplification pattern by the primers in the DRB5*01:08N kit^{1,2}.

¹DRB5 alleles listed on the IMGT/HLA web page 2017-April-13, release 3.28.0, www.ebi.ac.uk/imgt/hla.

²Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page http://hla.alleles.org/alleles/deleted.html.

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SPECIFICITY TABLE

DRB5*01:08N SSP subtyping

Specificities and sizes of the PCR products of the 2+1 primer mixes used for DRB5*01:08N SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control Amplified DRB5 alleles ³ band ²	
1	195 bp	515 bp	*01:08N
2 ⁴	175 bp	430 bp	*01:01:01-01:31, 02:02-02:16
3 ⁵	-	-	Negative Control

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DRB5*01:08N SSP typings. Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers. PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherit feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³For several DRB alleles 1st and/or 3rd exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

⁴In primer mix 2 the positive control band may be weaker than for other DRB5*01:08N primer mixes.

⁵Primer mix 3 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.

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Lot No.: **5F6** Lot-specific information

PRIMER SPECIFICATION

Well No.	1	2
Length of spec.	195	175
PCR product		
Length of int.	515	430
pos. control ¹		
5'-primer(s) ²	108(409)	13(125)
	^{5'} -AgA ^{3'}	^{5'} -gTA ^{3'}
3'-primer(s) ³	160(565)	57(258)
	5' -CAT 3'	^{5'} -gCg ^{3'}
		58(260)
		^{5'} -CCT ^{3'}
Well No.	1	2

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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CELL LINE VALIDATION SHEET						
DRB5*01:08N SSP kit ²						
					Well	
					1	2
					_	2
				<u>o</u> ::	2	70;
				Z	374	374
				Prod. No.:	201674701	201674702
		<u> </u>	DDDC	ш.	N	(1
		/C cell line ¹	DRB5			
2	9001	LK707	*04.02		-	-
3		E4181324	*01:02 *01:02		-	+
4		GU373	01.02		-	_
5		KAS011	*02:02		-	+
6	9353		02.02			_
7	9020				ļ <u>.</u>	
8	9020				ļ <u>.</u>	
9		YAR			-	
10		LKT3			-	-
11		PITOUT			 -	-
12	9052				-	-
13		JESTHOM			-	-
14		OLGA			-	-
15	9075				-	-
16		SWEIG007			-	-
17		CTM3953540			-	-
18		32367			-	-
19		BM16			-	-
20	9059	SLE005			-	-
21		AMALA			-	-
22	9056	KOSE			-	-
23	9124	IHL			-	-
24	9035	JBUSH			-	-
25	9049	IBW9			-	-
26	9285	WT49			-	-
27	9191	CH1007			-	-
28	9320	BEL5GB			-	-
29	9050	MOU			-	-
30	9021	RSH			-	-
31	9019	DUCAF			-	-
32	9297	HAG			-	-
33	9098	MT14B			-	-
34	9104	DHIF			-	-
35	9302	SSTO			-	-
36	9024	KT17			-	-
37		HHKB			-	-
38	9099	LZL			-	-
39	9315	CML			-	-
40	9134	WHONP199			_	-
41	9055	H0301			-	-
42	9066	TAB089			-	-
43	9076	T7526			-	-
44	9057	TEM			-	-
45	9239	SHJO			-	-
46	9013	SCHU	*01:01		-	+
47		TUBO			-	-
48	9303	TER-ND			_	-

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¹The provided cell line HLA specificities are retrieved from the http://www.ihwg.org/hla web site. The specificity of an individual cell line may thus be subject to change.

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

■LERUP SSP*

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